

# QuikChange<sup>®</sup> II XL Site-Directed Mutagenesis Kit

## INSTRUCTION MANUAL

Catalog #200521 (10 reactions) and #200522 (30 reactions)

Revision #063001b

**For In Vitro Use Only**



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# QuikChange® II XL Site-Directed Mutagenesis Kit

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# QuikChange® II XL Site-Directed Mutagenesis Kit

## MATERIALS PROVIDED

Materials provided	Quantity	
	Catalog #200522 <sup>a</sup> 30 reactions	Catalog #200521 <sup>b</sup> 10 reactions
<i>PfuUltra</i> ™ High Fidelity DNA polymerase (2.5 U/μl)	80 U	25 U
10× reaction buffer	500 μl	500 μl
<i>Dpn</i> I restriction enzyme (10 U/μl)	300 U	100 U
Oligonucleotide control primer #1 [34-mer (100 ng/μl)] 5' CCA TGA TTA CGC CAA GCG CGC AAT TAA CCC TCA C 3'	750 ng	750 ng
Oligonucleotide control primer #2 [34-mer (100 ng/μl)] 5' GTG AGG GTT AAT TGC GCG CTT GGC GTA ATC ATG G 3'	750 ng	750 ng
pWhitescript™ 4.5-kb control plasmid (5 ng/μl)	50 ng	50 ng
QuikSolution™ reagent	500 μl	500 μl
dNTP mix <sup>c</sup>	30 μl	10 μl
XL10-Gold® ultracompetent cells <sup>d</sup> (yellow tubes)	10 × 135 μl	4 × 135 μl
XL10-Gold® β-mercaptoethanol mix (β-ME)	2 × 50 μl	50 μl
pUC18 control plasmid (0.1 ng/μl in TE buffer <sup>e</sup> )	10 μl	10 μl

<sup>a</sup> The QuikChange II XL Site-Directed Mutagenesis Kit (Catalog #200522) contains enough reagents for 30 total reactions, which includes 5 control reactions.

<sup>b</sup> The QuikChange II XL Site-Directed Mutagenesis Kit (Catalog #200521) contains enough reagents for 10 total reactions, which includes 5 control reactions.

<sup>c</sup> Thaw the dNTP mix once, prepare single-use aliquots, and store the aliquots at –20°C. **Do not subject the dNTP mix to multiple freeze-thaw cycles.**

<sup>d</sup> Genotype: Tet<sup>r</sup>Δ (*mcrA*)183 Δ(*mcrCB-hsdSMR-mrr*)173 *endA1 supE44 thi-1 recA1 gyrA96 relA1 lac* Hte [F' *proAB lacZΔM15 Tn10* (Tet<sup>r</sup>) Amy Cam<sup>r</sup>]

<sup>e</sup> See *Preparation of Media and Reagents*.

## STORAGE CONDITIONS

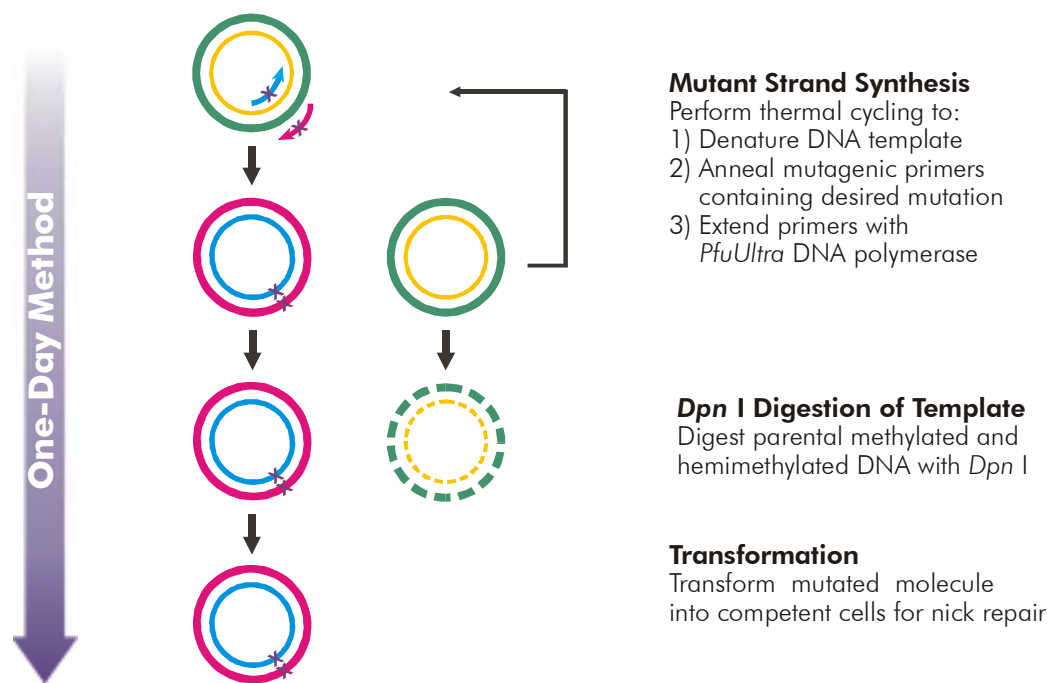
**XL10-Gold® Ultracompetent cells, XL10-Gold® β-ME, and pUC18 Control Plasmid: –80°C**  
**All Other Components: –20°C**

## ADDITIONAL MATERIALS REQUIRED

Falcon® 2059 polypropylene tubes (15 ml)  
5-Bromo-4-chloro-3-indoyl-β-D-galactopyranoside (X-gal)  
Isopropyl-1-thio-β-D-galactopyranoside (IPTG)

## INTRODUCTION

In vitro site-directed mutagenesis is an invaluable technique for characterizing the dynamic, complex relationships between protein structure and function, for studying gene expression elements, and for carrying out vector modification. Several approaches to this technique have been published, but these methods generally require single-stranded DNA (ssDNA) as the template<sup>1-4</sup> and are labor intensive or technically difficult. Stratagene's QuikChange® II XL site-directed mutagenesis kit\* is specifically optimized for large, difficult constructs and allows site-specific mutation in virtually any double-stranded plasmid. The kit eliminates the need for subcloning and for ssDNA rescue.<sup>5</sup> In addition, the QuikChange II XL site-directed mutagenesis kit does not require specialized vectors, unique restriction sites, multiple transformations or in vitro methylation treatment steps. The rapid three-step procedure generates mutants with greater than 80% efficiency (see Figure 1). The protocol is simple and uses either miniprep plasmid DNA or cesium-chloride-purified DNA.



**FIGURE 1** Overview of the QuikChange® II XL site-directed mutagenesis method.

\* U.S. Patent Nos. 5,789,166, 5,923,419, 6,391,548 and patents pending.

The QuikChange II XL kit is used to make point mutations, replace amino acids, and delete or insert single or multiple adjacent amino acids. The QuikChange II XL site-directed mutagenesis method is performed using *PfuUltra*<sup>™</sup> high-fidelity (HF) DNA polymerase\*\* for mutagenic primer-directed replication of both plasmid strands with the highest fidelity.<sup>¶</sup> The basic procedure utilizes a supercoiled double-stranded DNA (dsDNA) vector with an insert of interest and two synthetic oligonucleotide primers, both containing the desired mutation (see Figure 1). The oligonucleotide primers, each complementary to opposite strands of the vector, are extended during temperature cycling by *PfuUltra* HF DNA polymerase, without primer displacement. Extension of the oligonucleotide primers generates a mutated plasmid containing staggered nicks. Following temperature cycling, the product is treated with *Dpn* I. The *Dpn* I endonuclease (target sequence: 5'-Gm<sup>6</sup>ATC-3') is specific for methylated and hemimethylated DNA and is used to digest the parental DNA template and to select for mutation-containing synthesized DNA.<sup>¶</sup> DNA isolated from almost all *E. coli* strains is dam methylated and therefore susceptible to *Dpn* I digestion. The nicked vector DNA incorporating the desired mutations is then transformed into XL10-Gold<sup>®\*\*\*</sup> ultracompetent cells.

**Note** *While plasmid DNA isolated from almost all of the commonly used E. coli strains (dam<sup>+</sup>) is methylated and is a suitable template for mutagenesis, plasmid DNA isolated from the exceptional dam<sup>-</sup> E. coli strains, including JM110 and SCS110, is not suitable.*

The small amount of starting DNA template required to perform this method, the high fidelity of the *PfuUltra* HF DNA polymerase, and the low number of thermal cycles all contribute to the high mutation efficiency and decreased potential for random mutations during the reaction.

Stratagene's QuikChange II XL site-directed mutagenesis kit is derived from the QuikChange II site-directed mutagenesis method. The XL version of the kit is specialized for efficient mutagenesis of large or otherwise difficult-to-mutagenize plasmid templates and features components specifically designed for more efficient DNA replication and bacterial transformation. The QuikSolution<sup>™</sup> reagent is provided to facilitate replication of large plasmids, while XL10-Gold ultracompetent cells have been included to ensure the highest transformation efficiencies possible. The transformation efficiency of XL10-Gold cells is 5-fold higher than the transformation efficiency of XL1-Blue cells employed in the original QuikChange kit.<sup>7</sup> Moreover, XL10-Gold cells contain the Hte phenotype, which increases the transformation efficiency of larger DNA plasmids.

<sup>¶</sup> *PfuUltra* HF DNA polymerase has 18-fold higher fidelity in DNA synthesis than *Taq* DNA polymerase.

\*\* U.S. Patent Nos. 6,183,997, 6,333,165, 6,379,553, 5,948,663, 5,866,395, 5,545,552 and Patents Pending

\*\*\*U.S. Patent Nos. 5,512,486 and 5,707,841 and Patents Pending.

## QUIKCHANGE® II XL MUTAGENESIS CONTROL

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To demonstrate the effectiveness of the QuikChange II XL method, the pWhitescript™ 4.5-kb control plasmid is used to test the efficiency of mutant plasmid generation. The pWhitescript 4.5-kb control plasmid contains a stop codon (TAA) at the position where a glutamine codon (CAA) would normally appear in the  $\beta$ -galactosidase gene of the pBluescript® II SK(-) phagemid (corresponding to amino acid 9 of the protein). XL10-Gold ultracompetent cells transformed with this control plasmid appear white on LB-ampicillin agar plates (see *Preparation of Media and Reagents*), containing IPTG and X-gal, because  $\beta$ -galactosidase activity has been obliterated. The oligonucleotide control primers create a point mutation that reverts the T residue of the stop codon (TAA) in the  $\beta$ -galactosidase gene encoded on the pWhitescript 4.5-kb control template to a C residue to produce a glutamine codon (Gln, CAA). Following transformation, colonies can be screened for  $\beta$ -galactosidase production ( $\beta$ -gal<sup>+</sup>) by virtue of a blue colony phenotype.

# MUTAGENIC PRIMER DESIGN

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## Primer Design Guidelines

Stratagene has developed a web-based primer design software program specifically for designing optimal mutagenic primers for use with the QuikChange II XL site-directed mutagenesis kit. This program incorporates the design guidelines listed below, as well as additional primer design parameters involving free energy, mismatches, and codon replacement strategy. Go to <http://labtools.stratagene.com/QC> to design template-specific mutagenic primers using this program.

The mutagenic oligonucleotide primers for used with this protocol must be designed individually according to the desired mutation. The following considerations should be made for designing mutagenic primers:

- ◆ Both mutagenic primers must contain the desired mutation and anneal to the same sequence on opposite strands of the plasmid.
- ◆ Primers should be between 25 and 45 bases in length, and the melting temperature ( $T_m$ ) of the primers should be greater than or equal to 78°C. The following formula is commonly used for estimating the  $T_m$  of primers:

$$T_m = 81.5 + 0.41(\%GC) - 675/N - \% \text{ mismatch}$$

For calculating  $T_m$ :

- $N$  is the primer length in bases.
- values for **%GC** and **% mismatch** are whole numbers

For calculating  $T_m$  for primers intended to introduce insertions or deletions, use this modified version of the above formula:

$$T_m = 81.5 + 0.41(\%GC) - 675/N$$

where  $N$  does not include the bases which are being inserted or deleted.

- ◆ The desired mutation (deletion or insertion) should be in the middle of the primer with ~10–15 bases of correct sequence on both sides.
- ◆ The primers optimally should have a minimum GC content of 40% and should terminate in one or more C or G bases.

## Additional Primer Considerations

- ◆ Primers need not be 5' phosphorylated but **must** be purified either by fast polynucleotide liquid chromatography (FPLC) or by polyacrylamide gel electrophoresis (PAGE). Failure to purify the primers results in a significant decrease in mutation efficiency.
- ◆ It is important to keep primer concentration in excess. Stratagene suggests varying the amount of template while keeping the concentration of the primer constantly in excess.

## XL10-GOLD® ULTRACOMPETENT CELLS

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Stratagene's XL10-Gold ultracompetent cells, a derivative of Stratagene's highest-efficiency competent cell line XL2-Blue MRF', possess the Hte phenotype, which increases transformation efficiency of ligated DNA.<sup>7</sup> XL10-Gold cells are both endonuclease deficient (*endA1*) and recombination deficient (*recA*). The *endA1* mutation greatly improves the quality of plasmid miniprep DNA,<sup>8</sup> and the *recA* mutation helps ensure insert stability. In addition, the McrA, McrCB, McrF, Mrr, and HsdR systems have been removed from XL10-Gold ultracompetent cells. The *mcrA*, *mcrCB* and *mrr* mutations prevent cleavage of cloned DNA that carries cytosine and/or adenine methylation, which is often present in eukaryotic DNA and cDNA.<sup>9-11</sup> The McrA and McrCB systems recognize and restrict methylated cytosine DNA sequences, and the Mrr system recognizes and restricts methylated adenine DNA sequences. The Mrr system also restricts methylated cytosine DNA sequences with a specificity differing from that of McrA and McrCB. This activity has been named McrF. This McrF activity against methylated cytosines has been shown to be equal to or greater than the restriction activity of the McrA and McrCB systems.<sup>12</sup> The *hsdR* mutation prevents the cleavage of cloned DNA by the *EcoK* (*hsdR*) endonuclease system. XL10-Gold cells grow faster than XL1 or XL2-Blue cells, resulting in larger colonies. To permit blue-white color screening, the XL10-Gold ultracompetent cells contain the *lacI<sup>q</sup>ZΔM15* gene on the F' episome.

Host strain	References	Genotype
XL10-Gold® ultracompetent cells	7, 13, 14	Tet <sup>R</sup> Δ( <i>mcrA</i> )183 Δ( <i>mcrCB</i> - <i>hsdSMR</i> - <i>mrr</i> )173 <i>endA1 supE44 thi-1 recA1 gyrA96 relA1 lac Hte</i> [F' <i>proAB lacI<sup>q</sup>ZΔM15</i> Tn10 (Tet <sup>R</sup> ) Amy Cam <sup>R</sup> ] <sup>o</sup>

<sup>o</sup> Chloramphenicol resistant (Cam<sup>R</sup>) at concentrations of <40 µg/ml, but chloramphenicol sensitive (Cam<sup>S</sup>) at concentrations of 100 µg/ml.

It is important to store the XL10-Gold ultracompetent cells at -80°C to prevent a loss of efficiency. For best results, please follow the directions outlined in the following sections.

## PROTOCOL

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### QuikSolution™ Reagent

QuikSolution reagent has been shown to improve linear amplification. Enhanced amplification efficiencies are observed when using between 2.5–3.5 µl QuikSolution/50 µl reaction, with 3 µl being optimal for most targets.

### Mutant Strand Synthesis Reaction (Thermal Cycling)

**Notes** *Ensure that the plasmid DNA template is isolated from a dam<sup>+</sup> E. coli strain. The majority of the commonly used E. coli strains are dam<sup>+</sup>. Plasmid DNA isolated from dam<sup>-</sup> strains (e.g. JM110 and SCS110) is not suitable.*

*To maximize temperature-cycling performance, Stratagene strongly recommends using thin-walled tubes, which ensure ideal contact with the temperature cycler's heat blocks. The following protocols were optimized using thin-walled tubes.*

1. Synthesize two complimentary oligonucleotides containing the desired mutation, flanked by unmodified nucleotide sequence. Purify these oligonucleotide primers prior to use in the following steps (see *Mutagenic Primer Design*).
2. Prepare the control reaction as indicated below:

5 µl of 10× reaction buffer  
2 µl (10 ng) of pWhitescript 4.5-kb control plasmid (5 ng/µl)  
1.25 µl (125 ng) of oligonucleotide control primer #1  
    [34-mer (100 ng/µl)]  
1.25 µl (125 ng) of oligonucleotide control primer #2  
    [34-mer (100 ng/µl)]  
1 µl of dNTP mix  
3 µl of QuikSolution reagent  
36.5 µl of double-distilled water (ddH<sub>2</sub>O) to a final volume of 50 µl

Then add

1 µl of *PfuUltra* HF DNA polymerase (2.5 U/µl)

- Prepare the sample reaction(s) as indicated below:

**Note** *Stratagene recommends setting up an initial sample reaction using 10 ng of dsDNA template. If this initial reaction is unsuccessful, set up a series of sample reactions using various concentrations of dsDNA template ranging from 5 to 50 ng (e.g., 5, 10, 20, and 50 ng of dsDNA template) while keeping the primer concentration constant.*

5  $\mu$ l of 10 $\times$  reaction buffer  
 X  $\mu$ l (10 ng) of dsDNA template  
 X  $\mu$ l (125 ng) of oligonucleotide primer #1  
 X  $\mu$ l (125 ng) of oligonucleotide primer #2  
 1  $\mu$ l of dNTP mix  
 3  $\mu$ l of QuikSolution  
 ddH<sub>2</sub>O to a final volume of 50  $\mu$ l

Then add

1  $\mu$ l of *PfuUltra* HF DNA polymerase (2.5 U/ $\mu$ l)

- If the thermal cycler to be used does not have a hot-top assembly, overlay each reaction with ~30  $\mu$ l of mineral oil.
- Cycle each reaction using the cycling parameters outlined in Table I. (For the control reaction, use a 5-minute extension time and run the reaction for 12 cycles.)

**Note** *It is important to adhere to the 18-cycle limit when cycling the mutagenesis reactions. More than 18 cycles can have deleterious effects on the reaction efficiency.*

- Following temperature cycling, place the reaction tubes on ice for 2 minutes to cool the reactions to  $\leq 37^{\circ}\text{C}$ .

**Note** *If desired, amplification may be checked by electrophoresis of 10  $\mu$ l of the product on a 1% agarose gel. A band may or may not be visualized at this stage. In either case proceed with Dpn I digestion and transformation.*

**TABLE I**

**Cycling Parameters for the QuikChange® II XL Method**

Segment	Cycles	Temperature	Time
1	1	95°C	1 minute
2	18	95°C	50 seconds
		60°C	50 seconds
		68°C	1 minute/kb of plasmid length
3	1	68°C	7 minutes

\* For example, a 5-kb plasmid requires 5 minutes at 68°C per cycle.

## Dpn I Digestion of the Amplification Products

**Note** *It is important to insert the pipet tip below the mineral oil overlay (if used) when adding the Dpn I restriction enzyme to the reaction tubes during the digestion step and when removing the 1  $\mu$ l of the Dpn I-treated DNA for transfer to the transformation reaction. Stratagene suggests using specialized aerosol-resistant pipet tips, which are small and pointed, to facilitate this process.*

1. Add 1  $\mu$ l of the Dpn I restriction enzyme (10 U/ $\mu$ l) directly to each amplification reaction below the mineral oil overlay using a small, pointed pipet tip.
2. Gently and thoroughly mix each reaction mixture by pipetting the solution up and down several times. Spin down the reaction mixtures in a microcentrifuge for 1 minute, then immediately incubate the reactions at 37°C for 1 hour to digest the parental (i.e., the nonmutated) supercoiled dsDNA.

## Transformation of XL10-Gold® Ultracompetent Cells

**Notes** *Please read the Transformation Guidelines before proceeding with the transformation protocol.*

*XL10-Gold cells are resistant to tetracycline. If the mutagenized plasmid contains only the tet<sup>R</sup> resistance marker, an alternative tetracycline-sensitive strain of competent cells must be used.*

1. Gently thaw the XL10-Gold ultracompetent cells on ice. For each control and sample reaction to be transformed, aliquot 45  $\mu$ l of the ultracompetent cells to a *prechilled* Falcon® 2059 polypropylene tube.
2. Add 2  $\mu$ l of the  $\beta$ -ME mix provided with the kit to the 45  $\mu$ l of cells. (Stratagene cannot guarantee highest efficiencies with  $\beta$ -ME from other sources.)
3. Swirl the contents of the tube gently. Incubate the cells on ice for 10 minutes, swirling gently every 2 minutes.
4. Transfer 2  $\mu$ l of the Dpn I-treated DNA from each control and sample reaction to separate aliquots of the ultracompetent cells.

**Note** *Carefully remove any residual mineral oil from the pipet tip before transferring the Dpn I-treated DNA to the transformation reaction.*

As an optional control, verify the transformation efficiency of the XL10-Gold ultracompetent cells by adding 1  $\mu\text{l}$  of 0.01 ng/ $\mu\text{l}$  pUC18 control plasmid (dilute the control provided 1:10 in high-quality water) to another 45- $\mu\text{l}$  aliquot of cells.

Swirl the transformation reactions gently to mix and incubate the reactions on ice for 30 minutes.

5. Preheat NZY<sup>+</sup> broth (see *Preparation of Media and Reagents*) in a 42°C water bath for use in step 8.

**Note** *Transformation of XL10-Gold ultracompetent cells has been optimized using NZY<sup>+</sup> broth.*

6. Heat-pulse the tubes in a 42°C water bath for 30 seconds. The duration of the heat pulse is *critical* for obtaining the highest efficiencies. Do not exceed 42°C.

**Note** *This heat pulse has been optimized for transformation in Falcon 2059 polypropylene tubes.*

7. Incubate the tubes on ice for 2 minutes.
8. Add 0.5 ml of preheated (42°C) NZY<sup>+</sup> broth to each tube, then incubate the tubes at 37°C for 1 hour with shaking at 225–250 rpm.
9. Plate the appropriate volume of each transformation reaction, as indicated in the table below, on agar plates containing the appropriate antibiotic for the plasmid vector.

For the mutagenesis and transformation controls, spread cells on LB–ampicillin agar plates containing 80  $\mu\text{g}/\text{ml}$  X-gal and 20 mM IPTG (see *Preparing the Agar Plates for Color Screening*).

#### Transformation reaction plating volumes

Reaction Type	Volume to Plate
pWhitescript mutagenesis control	250 $\mu\text{l}$
pUC18 transformation control	5 $\mu\text{l}$ (in 200 $\mu\text{l}$ of NZY <sup>+</sup> broth)*
Sample mutagenesis	250 $\mu\text{l}$ on each of two plates (entire transformation reaction)

\* Place a 200- $\mu\text{l}$  pool of NZY<sup>+</sup> broth on the agar plate, pipet the 5  $\mu\text{l}$  of the transformation reaction into the pool, then spread the mixture.

10. Incubate the transformation plates at 37°C for >16 hours.

### Expected Results for the Control Transformations

The expected colony number from the transformation of the pWhitescript 4.5 kb control mutagenesis reaction is between 50 and 800 colonies. Greater than 80% of the colonies should contain the mutation and appear as blue colonies on agar plates containing IPTG and X-gal.

**Note** *The mutagenesis efficiency (ME) for the pWhitescript 4.5-kb control plasmid is calculated by the following formula:*

$$ME = \frac{\text{Number of blue colony forming units (cfu)}}{\text{Total number of colony forming units (cfu)}} \times 100\%$$

If transformation of the pUC18 control plasmid was performed, >100 colonies (>10<sup>9</sup> cfu/μg) should be observed, with >98% having the blue phenotype.

### Expected Results for Sample Transformations

The expected colony number is between 10 and 1000 colonies, depending upon the base composition and length of the DNA template employed. For suggestions on increasing colony number, see *Troubleshooting*. The insert of interest should be sequenced to verify that selected clones contain the desired mutation(s).

## TRANSFORMATION GUIDELINES

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### Storage Conditions

Ultracompetent cells are sensitive to even small variations in temperature and must be stored at the bottom of a  $-80^{\circ}\text{C}$  freezer. Transferring tubes from one freezer to another may result in a loss of efficiency. Ultracompetent cells should be placed at  $-80^{\circ}\text{C}$  directly from the dry ice shipping container.

### Aliquoting Cells

When aliquoting, keep ultracompetent cells on ice at all times. It is essential that the Falcon 2059 polypropylene tubes are placed on ice before the cells are thawed and that the cells are aliquoted directly into the prechilled tubes.

### Use of Falcon® 2059 Polypropylene Tubes

It is important that Falcon 2059 polypropylene tubes are used for the transformation protocol, since other tubes may be degraded by the  $\beta$ -mercaptoethanol used in the *Transformation Protocol*. In addition, the duration of the heat-pulse step is critical and has been optimized specifically for the thickness and shape of the Falcon 2059 polypropylene tubes.

### Use of $\beta$ -Mercaptoethanol

$\beta$ -Mercaptoethanol ( $\beta$ -ME) has been shown to increase transformation efficiency. The XL10-Gold  $\beta$ -mercaptoethanol mix provided in this kit is diluted and ready to use.

### Quantity of DNA Added

Greatest efficiencies are observed when adding 2  $\mu\text{l}$  of the ligation mixture. A greater number of colonies will be obtained when adding up to 50 ng, although the overall efficiency may be lower.

### Length and Temperature of the Heat Pulse

There is a defined window of highest efficiency resulting from the heat pulse during transformation. Optimal efficiencies are observed when cells are heat-pulsed for 30 seconds. Heat-pulsing for at least 30 seconds is recommended to allow for slight variations in the length of incubation. Efficiencies decrease when incubating for  $<30$  seconds or for  $>40$  seconds. Do not exceed  $42^{\circ}\text{C}$ .

### Preparing the Agar Plates for Color Screening

To prepare the LB agar plates for blue–white color screening, add 80  $\mu\text{g}/\text{ml}$  of 5-bromo-4-chloro-3-indolyl- $\beta$ -D-galactopyranoside (X-gal), 20 mM isopropyl-1-thio- $\beta$ -D-galactopyranoside (IPTG), and the appropriate antibiotic to the LB agar. Alternatively, 100  $\mu\text{l}$  of 10 mM IPTG and 100  $\mu\text{l}$  of 2% X-gal can be spread on the LB agar plates 30 minutes prior to plating the transformations. Prepare the IPTG in sterile  $\text{dH}_2\text{O}$ ; prepare the X-gal in dimethylformamide (DMF). Do not mix the IPTG and X-gal before pipetting them onto the plates because these chemicals may precipitate.

## TROUBLESHOOTING

When used according to the guidelines outlined in this instruction manual, Stratagene's kit will provide a reliable means to conduct site-directed mutagenesis using dsDNA templates. Variations in the base composition and length of the DNA template and in thermal cycler performance may contribute to differences in mutagenesis efficiency. Stratagene provides the following guidelines for troubleshooting these variations.

Observation	Suggestion(s)
Low transformation efficiency or low colony number	Ensure that sufficient DNA template is used in the reaction. Visualize the DNA template on a gel to verify the quantity and quality. Repeat reaction using higher amounts of plasmid DNA (100 ng, 200 ng, 500 ng)
	Ensure that sufficient mutant DNA is synthesized in the reaction. <ul style="list-style-type: none"> <li>• Titrate QuikSolution™ reagent in 1-<math>\mu</math>l increments from 0 to 5 <math>\mu</math>l</li> <li>• Increase the amount of the Dpn I-treated DNA used in the transformation reaction to 4 <math>\mu</math>l</li> <li>• Increase the extension time to 2.5 min/kb</li> <li>• Precipitate the entire reaction and use all of it in the transformation</li> </ul>
	Ensure sufficient mutant DNA is synthesized by adjusting the cycling parameters for the sample reaction to overcome differences in ramping efficiencies of thermal cyclers. Increase initial denaturation step (segment 1) to 1–2 minutes and denaturation cycles (segment 2) to 1 minute
	Ensure that excess mineral oil is not transferred into the transformation reaction when pipetting the Dpn I-treated DNA. Using the smallest pipet tips available, insert the pipet tip completely below the mineral layer overlay and clear the pipet tip while submerged beneath the mineral oil overlay before collecting the sample
Low mutagenesis efficiency or low colony number with the control reaction	Different thermal cyclers contribute to variations in cycling efficiencies. Optimize the cycling parameters (including ramp rates) for the control reaction then repeat the protocol for the sample reactions using the optimized conditions
	Ensure that ultracompetent cells are stored at the bottom of a –80°C freezer immediately upon arrival; use XL10-Gold $\beta$ -ME in the transformation reactions (see also <i>Transformation Guidelines</i> )
	Verify that the agar plates were prepared correctly. See <i>Preparing the Agar Plates for Color Screening</i> , and follow the recommendations for IPTG and X-Gal concentrations carefully
	For best visualization of the blue ( $\beta$ -gal <sup>+</sup> ) phenotype, the control plates must be incubated for at least 16 hours at 37°C
	Avoid multiple freeze-thaw cycles for the dNTP mix. Thaw the dNTP mix once, prepare single-use aliquots, and store the aliquots at –20°C. Do not subject the dNTP mix to multiple freeze-thaw cycles
Low mutagenesis efficiency with the sample reaction(s)	Add the Dpn I restriction enzyme below the mineral oil overlay in the digestion step and ensure proper mixing of all components, especially the Dpn I, in the reaction
	Allow sufficient time for the Dpn I to completely digest the parental template; repeat the digestion if too much DNA template was present. Increase digestion time to 1.5–2.0 hours
	Avoid multiple freeze-thaw cycles for the dNTP mix. Thaw the dNTP mix once, prepare single-use aliquots, and store the aliquots at –20°C. Do not subject the dNTP mix to multiple freeze-thaw cycles
False positives	Poor quality primers can lead to false positives. Radiolabel the primers and check for degradation on an acrylamide gel or resynthesize the primers
	False priming can lead to false positives. Increase the stringency of the reaction by increasing the annealing temperature to within 5°C of the melting temperature of the mutagenic primers

## PREPARATION OF MEDIA AND REAGENTS

<p><b>LB Agar (per Liter)</b> 10 g of NaCl 10 g of tryptone 5 g of yeast extract 20 g of agar Add deionized H<sub>2</sub>O to a final volume of 1 liter Adjust pH to 7.0 with 5 N NaOH Autoclave Pour into petri dishes (~25 ml/100-mm plate)</p>	<p><b>LB–Ampicillin Agar (per Liter)</b> 1 liter of LB agar Autoclave Cool to 55°C Add 100 mg of filter-sterilized ampicillin Pour into petri dishes (~25 ml/100-mm plate)</p>
<p><b>NZY<sup>+</sup> Broth (per Liter)</b> 10 g of NZ amine (casein hydrolysate) 5 g of yeast extract 5 g of NaCl Add deionized H<sub>2</sub>O to a final volume of 1 liter Adjust to pH 7.5 using NaOH Autoclave Add the following filter-sterilized supplements prior to use: 12.5 ml of 1 M MgCl<sub>2</sub> 12.5 ml of 1 M MgSO<sub>4</sub> 20 ml of 20% (w/v) glucose (or 10 ml of 2 M glucose)</p>	<p><b>TE Buffer</b> 10 mM Tris-HCl (pH 7.5) 1 mM EDTA</p>

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## ENDNOTES

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## QuikChange® II XL Site-Directed Mutagenesis Kit

Catalog #200521, #200522

### QUICK-REFERENCE PROTOCOL

- Prepare the control and sample reaction(s) as indicated below:

**Note** *Stratagene recommends setting up an initial sample reaction using 10 ng of dsDNA template. If this initial sample reaction is unsuccessful, set up a series of reactions using various amounts of dsDNA template ranging from 5 to 50 ng (e.g., 5, 10, 20, and 50 ng of dsDNA template) while keeping the primer concentration constant.*

#### Control Reaction

- 5  $\mu$ l of 10 $\times$  reaction buffer
- 2  $\mu$ l (10 ng) of pWhitescript™ 4.5-kb control template (5 ng/ $\mu$ l)
- 1.25  $\mu$ l (125 ng) of oligonucleotide control primer #1 [34-mer (100 ng/ $\mu$ l)]
- 1.25  $\mu$ l (125 ng) of oligonucleotide control primer #2 [34-mer (100 ng/ $\mu$ l)]
- 1  $\mu$ l of dNTP mix
- 3  $\mu$ l of QuikSolution™ reagent
- 35.5  $\mu$ l ddH<sub>2</sub>O to a final volume of 50  $\mu$ l

#### Sample Reaction

- 5  $\mu$ l of 10 $\times$  reaction buffer
- X  $\mu$ l (10 ng) of dsDNA template
- X  $\mu$ l (125 ng) of oligonucleotide primer #1
- X  $\mu$ l (125 ng) of oligonucleotide primer #2
- 1  $\mu$ l of dNTP mix
- 3  $\mu$ l of QuikSolution™ reagent
- ddH<sub>2</sub>O to a final volume of 50  $\mu$ l

- Then add 1  $\mu$ l of *PfuUltra* HF DNA polymerase (2.5 U/ $\mu$ l) to each control and sample reaction
- If the thermal cycler to be used does not have a hot top assembly, overlay each reaction with  $\sim$ 30  $\mu$ l of mineral oil.
- Cycle each reaction using the cycling parameters outlined in the following table:

Segment	Cycles	Temperature	Time
1	1	95°C	1 minute
2	18	95°C	50 seconds
		60°C	50 seconds
		68°C	1 minute/kb of plasmid length
3	1	68°C	7 minutes

- Add 1  $\mu$ l of *Dpn* I restriction enzyme (10 U/ $\mu$ l) below the mineral oil overlay
- Gently and thoroughly mix each reaction, spin down in a microcentrifuge for 1 minute, and immediately incubate at 37°C for 1 hour to digest the parental supercoiled dsDNA
- Transform 2  $\mu$ l of the *Dpn* I-treated DNA from each control and sample reaction into separate 45- $\mu$ l aliquots of XL10-Gold ultracompetent cells (see *Transformation of XL10-Gold® Ultracompetent Cells* in the instruction manual).